

This form should be used for all taxonomic proposals. Please complete all those modules that are applicable (and then delete the unwanted sections). For guidance, see the notes written in blue and the separate document "Help with completing a taxonomic proposal"

Please try to keep related proposals within a single document; you can copy the modules to create more than one genus within a new family, for example.

MODULE 1: TITLE, AUTHORS, etc

Code assigned:	2016.065a-dB			(to be completed by ICTV officers)				
Short title: To create one (1) not family <i>Siphoviridae</i> . (e.g. 6 new species in the genus 2 Modules attached (modules 1 and 10 are required)	12virus, ir 1 ⊠ 6 □	ncluding tw	3	w species in 4	5 ☐ 10 ⊠			
Author(s):								
Andrew M. Kropinski – Univer Evelien M. Adriaenssens – Uni	, ,	/	n Africa)					
Corresponding author with e	-mail address:							
Andrew M. Kropinski Phage.C	anada@gmail.o	<u>com</u>						
List the ICTV study group(s)	that have seen	n this pro	posal:					
A list of study groups and contacts http://www.ictvonline.org/subcomm in doubt, contact the appropriate s chair (fungal, invertebrate, plant, p vertebrate viruses)	mittees.asp . If subcommittee	Subcom	Bacterial amittee	l and	Archaeal	Viruses		
ICTV Study Group comments (if any) and response of the proposer:								
Date first submitted to ICTV: Date of this revision (if differen	, and a second s							
ICTV-EC comments and response of the proposer:								

MODULE 2: NEW SPECIES

creating and naming one or more new species.

If more than one, they should be a group of related species belonging to the same genus. All new species must be placed in a higher taxon. This is usually a genus although it is also permissible for species to be "unassigned" within a subfamily or family. Wherever possible, provide sequence accession number(s) for **one** isolate of each new species proposed.

Code	201	6.065aB	(assigned by IC	CTV officers)			
To crea	ate 2 no	ew species within:					
Genus: Ydn12virus (new) Subfamily: Family: Siphoviridae Order: Caudovirales				Fill in all that apply. If the higher taxon has yet to be created (in a later module, below) write "(new)" after its proposed name. If no genus is specified, enter "unassigned" in the genus box.			
Name	of new	species:	Representative species please)	isolate: (only 1 per	GenBank sequence accession number(s)		
Streptomyces virus YDN12 Streptomyces virus TP1604		Streptomyces phage YDN12 Streptomyces phage TP1604		KP876465.1 KP876466.1			

Reasons to justify the creation and assignment of the new species:

- Explain how the proposed species differ(s) from all existing species.
 - If species demarcation criteria (see module 3) have previously been defined for the genus, explain how the new species meet these criteria.
 - o If criteria for demarcating species need to be defined (because there will now be more than one species in the genus), please state the proposed criteria.
- Further material in support of this proposal may be presented in the Appendix, Module 9

We have chosen 95% DNA sequence identity as the criterion for demarcation of species in this new genus. Each of the proposed species differs from the others with more than 5% at the DNA level as confirmed with the BLASTN algorithm.

MODULE 3: NEW GENUS

creating a new genus

Ideally, a genus should be placed within a higher taxon.

Code	201	6.065bB	(assigned by ICTV officers)			
To create	a new	genus within:		Fill in all that apply.		
Subfa	mily:			If the higher taxon has yet to be created """ """ """ """ """ """ """		
Fa	mily:	Siphoviridae		(in a later module, below) write "(new)" after its proposed name.		
Order: Caudovirales			If no family is specified, enter "unassigned" in the family box			

naming a new genus

Code	2016.065cB	(assigned by ICTV officers)			
To name the new genus: Ydn12virus (new)					

Assigning the type species and other species to a new genus

Code	2016.065dB	(assigned by ICTV officers)						
To desig	To designate the following as the type species of the new genus							
Streptomyces virus YDN12 Every genus must have a type species. This should be a well characterized species although not necessarily the first to be discovered								
The new genus will also contain any other new species created and assigned to it (Module 2) and any that are being moved from elsewhere (Module 7b). Please enter here the TOTAL number of species (including the type species) that the genus will contain: 2								

Reasons to justify the creation of a new genus:

Additional material in support of this proposal may be presented in the Appendix, Module 9

Phage YDN12 was enriched from soil from Richardson (TX, USA) while TP1604 came from Keller (TX) on *Streptomyces griseus* ATCC 10137. Phages of this type have circularly permuted genomes. The Actinobacteriophage Database classifies these phages into Cluster BG (http://phagesdb.org/clusters/BG/).

BLASTN, CoreGenes (Table 1) [2], progressiveMauve alignment (Fig. 2) [1] and phylogenetic analyses (Fig. 3) [3] all indicate that the proposed genus, *Ynd12virus*, is cohesive and distinct from the other genera of viruses. On average the genomes of this genus are 56.85 kb (69.2 mol% G+C), and encode71 proteins and 0 tRNAs.

Origin of the new genus name:

Based upon the name of the first sequenced member of this genus

Reasons to justify the choice of type species:

The first sequenced member of this genus

Species demarcation criteria in the new genus:

If there will be more than one species in the new genus, list the criteria being used for species demarcation and explain how the proposed members meet these criteria.

We have chosen 95% DNA sequence identity as the criterion for demarcation of species in this new genus. Each of the proposed species differs from the others with more than 5% at the DNA level as confirmed with the BLASTN algorithm.

MODULE 10: APPENDIX: supporting material

additional material in support of this proposal

References:

- 1. Darling AE, Mau B, Perna NT. progressiveMauve: multiple genome alignment with gene gain, loss and rearrangement. PLoS One. 2010; 5(6):e11147.
- 2. Turner D, Reynolds D, Seto D, Mahadevan P. CoreGenes3.5: a webserver for the determination of core genes from sets of viral and small bacterial genomes. BMC Res Notes. 2013; 6:140. doi: 10.1186/1756-0500-6-140.
- 3. Dereeper A, Guignon V, Blanc G, Audic S, Buffet S, Chevenet F, Dufayard JF, Guindon S, Lefort V, Lescot M, Claverie JM, Gascuel O. Phylogeny.fr: robust phylogenetic analysis for the non-specialist. Nucleic Acids Res. 2008; 36(Web Server issue):W465-9.

Annex:

Include as much information as necessary to support the proposal, including diagrams comparing the old and new taxonomic orders. The use of Figures and Tables is strongly recommended but direct pasting of content from publications will require permission from the copyright holder together with appropriate acknowledgement as this proposal will be placed on a public web site. For phylogenetic analysis, try to provide a tree where branch length is related to genetic distance.

Fig. 1. Electron micrograph of negatively stained phage TP1604 (Obtained from the Actinobacteriophage Database; http://phagesdb.org/phages/TP1604/). Limited permission was granted by The Actinobacteriophages Database, funded by the Howard Hughes Medical Institute, to use this electron micrograph for this taxonomy proposal; it cannot be reused without permission of The Actinobacteriophages Database.

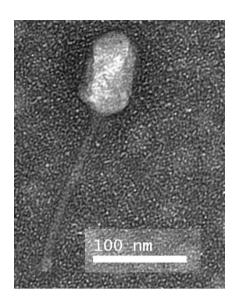


Table 1. Properties of the two phages belonging to the genus *Ydn12virus*.

Phage	RefSeq No.	GenBank	Genome	Genome	No.	DNA (%	%
		Accession	size	(mol%G+C)	CDS	sequence	Homologous
		No.	(kb)			identity)*	proteins **
YDN12	NC_028974	KP876465	56.53	69.2	71	100%	100%
TP1604	NC_028818	KP876466	57.17	69.2	71	87	91.4

^{*} Determined using BLASTN; ** Determined using CoreGenes [2]; *Streptomyces* phage Miah (KU189325) should be considered a strain in this genus.

Fig. 2. progressiveMauve alignment [1] of the annotated genomes of members of the *Ydn12virus* genus – from top to bottom: YDN12 and TP1604. Colored blocks indicate the regions of 1 to 1 best alignment with rearrangement breakpoints in a different random color. The degree of sequence similarity between regions is given by a similarity plot within the colored blocks with the height of the plot proportional to the average nucleotide identity (Aaron Darling, personal communication).

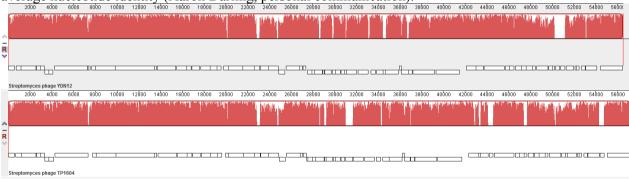


Fig. 3. Phylogenetic analysis of (A) large subunit terminase proteins and (B) major capsid proteins of YDN12-like viruses and homologous proteins from a variety of other phages constructed using "one click" at phylogeny.fr [3]. "The "One Click mode" targets users that do not wish to deal with program and parameter selection. By default, the pipeline is already set up to run and connect programs recognized for their accuracy and speed (MUSCLE for multiple alignment and PhyML for phylogeny) to reconstruct a robust phylogenetic tree from a set of sequences." It also includes the use of Gblocks to eliminate poorly

aligned positions and divergent regions. "The usual bootstrapping procedure is replaced by a new confidence index that is much faster to compute. See: Anisimova M., Gascuel O. Approximate likelihood ratio test for branches: A fast, accurate and powerful alternative (Syst Biol. 2006;55(4):539-52.) for details."

A. TerL proteins

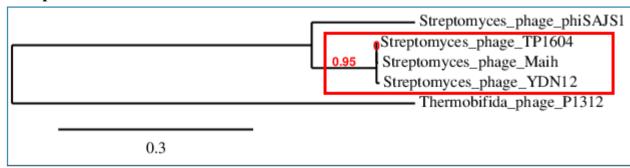


Figure 1: Phylogenetic tree.

B. Major capsid proteins

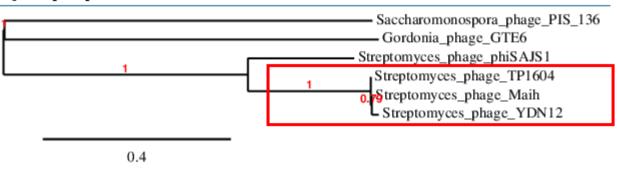


Figure 1: Phylogenetic tree.